

## Rapid Total Particle Quantification for Viral Families With High Potential for Pathogenic Emergence

Rebecca K. Montange, Ph.D., Katherine D. Shives, Ph.D., Jeffrey W. Steaffens, M.S., Mark S. Smith, M.S.  
Sartorius Virus Analytics, Arvada, CO

Rapid viral quantification during the emergence of a novel pathogenic virus is critical to quickly developing a vaccine. Current techniques for determining the quantity of virus particles in a sample, such as immunoassays, the plaque assay, TCID<sub>50</sub>, or measures of genomic content such as qPCR require a certain prior knowledge of the virus so that the correct cell lines can be utilized and the correct reagents manufactured. Building the suite of specific reagents for molecular or immunological studies can take weeks to months, with potentially devastating consequences during an emerging virus outbreak.

Here we demonstrate the capabilities of a novel technology for generalized and rapid virus particle quantification: the Virus Counter® 3100 (VC 3100) instrument, coupled with a two-component fluorogenic dye reagent designed to stain and allow detection of viral protein and nucleic acid, to directly quantify the total particle count of enveloped viral samples. This platform operates without the need for species- or strain-specific reagents. For this study, representative members from the Arenaviridae, Coronaviridae, Flaviviridae, Orthomyxoviridae, and Togaviridae families were utilized. Titer results for each virus sample were obtained using the Virus Counter® platform and compared to the titer data from the suppliers' Certificates of Analysis. These results demonstrate the efficacy of the Virus Counter® 3100 platform for rapid and precise quantification of viruses from families with high potential for evolving or harboring new human pathogens. This technology can therefore improve time-to-result for critical experiments in vaccine manufacturing that require viral particle counts for a novel virus.

### Staining Protocol for Virus Samples

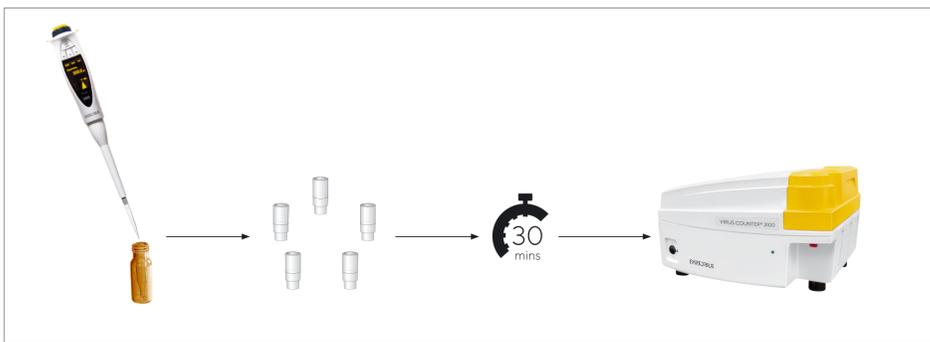


Figure 1

Diluted samples are mixed with a two-component fluorogenic dye reagent and incubated for 30 minutes (Figure 1). Stained samples are analyzed using the Virus Counter® 3100 instrument. Run time is three minutes per sample. Using similar principles to flow cytometry, intact particles are quantified and results reported as vp/mL.

### Serial Dilutions of Viral Strains

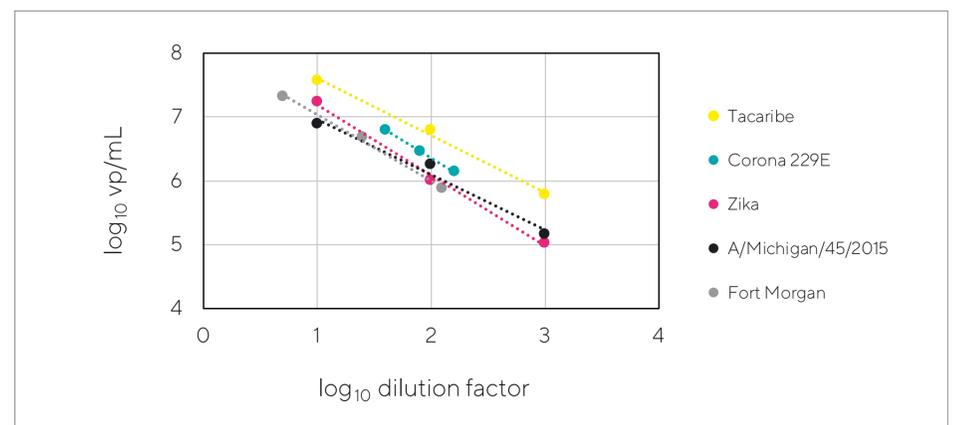


Figure 3

Serial dilutions of representative species from families with high potential for emergence as human pathogens (Figure 3). Our generalized reagent enables rapid and specific detection of a wide variety of enveloped viral species. While the influenza strain shown here is a seasonal flu strain, this technology can be used to detect subtypes with pandemic potential.

### Virus Particle Detection

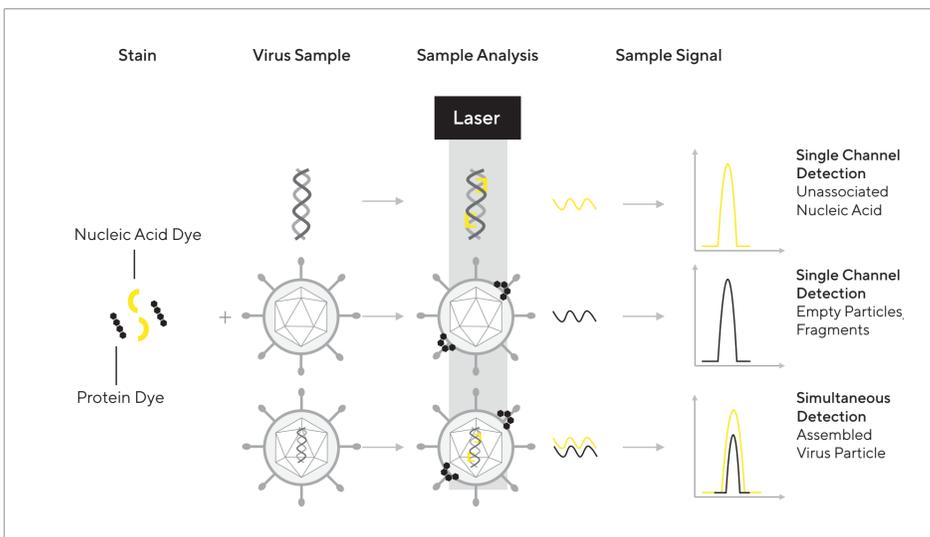


Figure 2

The two fluorogenic dye components are generalized stains for nucleic acid content and the hydrophobic regions of proteins. As these bound dyes flow through a laser beam, the emitted signals are detected on separate channels. The simultaneous signals are recorded as results (Figure 2).

Family	Species	Strain	VC 3100 Titer (vp/mL)	TCID <sub>50</sub> (TCID <sub>50</sub> /mL)*	Total   infectious particles**
Arenavirus	Tacaribe	TRVL-11573	1.24×10 <sup>8</sup> ± 5.60×10 <sup>7</sup>	8.6×10 <sup>6</sup>	14 (7.6 - 20.0)
Coronavirus	Human Corona	229E	4.70×10 <sup>8</sup> ± 1.31×10 <sup>8</sup>	1.58×10 <sup>6</sup>	297 (214 - 380)
Flavivirus	Zika	PRVABC59	2.20×10 <sup>8</sup> ± 9.94×10 <sup>7</sup>	2.8×10 <sup>7</sup>	7.8 (4.3 - 11)
Orthomyxovirus	Influenza A	A/Michigan/45/2015	8.19×10 <sup>7</sup> ± 1.68×10 <sup>7</sup>	1.6×10 <sup>7</sup>	5.1 (4.1 - 6.2)
Togavirus	Fort Morgan	CM4-146	1.29×10 <sup>8</sup> ± 4.38×10 <sup>7</sup>	1.58×10 <sup>6</sup>	0.815 (0.537 - 1.09)

\* TCID<sub>50</sub> data provided by vendor  
\*\* Range is 95% confidence interval for Virus Counter® data

Table 1

Tabulated comparison of VC 3100 results to infectious titer data with the total particle to infectious particle ratio (Table 1). The data was taken on crude samples.

### Comparison of Total Particle vs. Infectious Titers

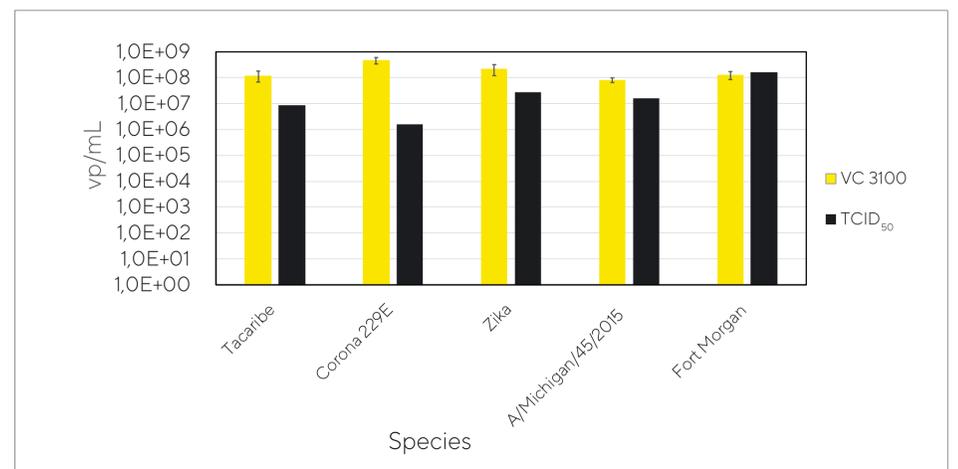


Figure 4

Titer comparisons of the VC 3100 results and infectious titer. VC 3100 results are shown with error bars representing the 95% confidence interval (Figure 4). The TCID<sub>50</sub> results were provided by the sample vendors and experimental error information was not available.

### Conclusions

- The Virus Counter® 3100 Platform enables rapid quantification of a variety of enveloped viral species.
- Titer results for virus samples are available in less than one hour.
- Rapid method to quantify total virus particles, including the non-infectious population.
- No prior knowledge of genomic sequences or envelope epitopes is required.

This platform can therefore improve time-to-result for critical experiments in vaccine manufacturing that require viral particle counts where other methods may not be readily available for a novel virus